

Genome sequence of *Ensifer medicae* strain WSM1115; an acid-tolerant *Medicago*-nodulating microsymbiont from Samothraki, Greece

Wayne Reeve^{1*}, Ross Ballard², John Howieson¹, Elizabeth Drew², Rui Tian¹, Lambert Bräu³, Christine Munk⁴, Karen Davenport⁴, Patrick Chain⁴, Lynne Goodwin⁴, Ioanna Pagani⁵, Marcel Huntemann⁵, Konstantinos Mavrommatis⁶, Amrita Pati⁵, Victor Markowitz⁶, Natalia Ivanova⁵, Tanja Woyke⁵ & Nikos Kyrpides⁵.

¹ Centre for Rhizobium Studies, Murdoch University, Western Australia, Australia

² South Australian Research and Development Institute, Urrbrae, South Australia, Australia

³ School of Life and Environmental Sciences, Deakin University, Victoria, Australia

⁴ Los Alamos National Laboratory, Bioscience Division, Los Alamos, New Mexico, USA

⁵ DOE Joint Genome Institute, Walnut Creek, California, USA

⁶ Biological Data Management and Technology Center, Lawrence Berkeley National Laboratory, Berkeley, California, USA

*Correspondence: Wayne Reeve (W.Reeve@murdoch.edu.au)

Keywords: root-nodule bacteria, nitrogen fixation, rhizobia, *Alphaproteobacteria*

Ensifer medicae strain WSM1115 forms effective nitrogen fixing symbioses with a range of annual *Medicago* species and is used in commercial inoculants in Australia. WSM1115 is an aerobic, motile, Gram-negative, non-spore-forming rod. It was isolated from a nodule recovered from the root of burr medic (*Medicago polymorpha*) collected on the Greek Island of Samothraki. WSM1115 has a broad host range for nodulation and N fixation capacity within the genus *Medicago*, although this does not extend to all medic species. WSM1115 is considered saprophytically competent in moderately acid soils (pH(CaCl) 5.0), but it has failed to persist at field sites where soil salinity exceeded 10 ECe (dS/m). Here we describe the features of *E. medicae* strain WSM1115, together with genome sequence information and its annotation. The 6,861,065 bp high-quality-draft genome is arranged into 7 scaffolds of 28 contigs, contains 6,789 protein-coding genes and 83 RNA-only encoding genes, and is one of 100 rhizobial genomes sequenced as part of the DOE Joint Genome Institute 2010 Genomic Encyclopedia for *Bacteria* and *Archaea*-Root Nodule *Bacteria* (GEBA-RNB) project.

Introduction

The genus *Medicago* comprises 87 species of annual and perennial legumes, including some that were formerly recognized as *Trigonella* and *Melilotus* species [1]. A small number of annual *Medicago* species that have been domesticated are grown extensively in the sheep-wheat zone of southern Australia, particularly where pasture regeneration after a cropping phase is desirable. Annual *Medicago* species are grown on more than 20 M ha [2] and are particularly valued for their contribution to farming systems, in which *Medicago* fix around 25 kg of N per tonne of legume dry matter produced [3].

Medicago are nodulated by two species of root nodule bacteria (*Ensifer medicae* and *Ensifer meliloti*) that are recognized as being distinct based on their different nodulation and N₂ fixation

phenotypes in host interaction studies and more detailed analyses of their genetics [4,5].

Ensifer medicae strain WSM1115 is used in Australia to produce commercial peat cultures (referred to as Group AM inoculants) for the inoculation of several species of annual *Medicago* (predominantly *M. truncatula*, *M. polymorpha*, *M. scutellata*, *M. sphaerocarpos*, *M. murex*, *M. rugosa* and *M. orbicularis*). WSM1115 has been used commercially since 2002 [6], when it replaced strain WSM688. WSM1115 was isolated from a nodule from the roots of burr medic (*Medicago polymorpha*) collected by Prof. John Howieson (Murdoch University, Australia) on the island of Samothraki, Greece.

WSM1115 was selected for use in commercial inoculants having demonstrated good N₂-fixation capacity with the relevant medic hosts and adequate

saprophytic competence in moderately acidic soil (pH(CaCl₂) 5).

Saprophytic competence in acidic soils is a requirement of strains used to inoculate *Medicago* because several species (*M. murex*, *M. sphaerocarpus* and *M. polymorpha*) are recommended and sown into soils below pH(CaCl₂) 5.5, a level that is known to limit both survival of medic rhizobia and nodulation processes [7-10]. Useful variation in saprophytic competence occurs between strains of medic rhizobia [9] and valuable insights into the mechanisms that confer acidity tolerance have been provided by studies using strain WSM419 [11], which has been recently sequenced [12]. However, the complex nature of soil adaptation means that *in-situ* field studies still provide the most reliable means of selecting an inoculant strain and were used to select WSM1115 for commercial use. In a cross row experiment comparing 15 strains on acidic sand (pH(CaCl₂) 5.0; Dowerin, West Australia), the nodulation of plants inoculated with WSM1115 was equal to or better than that of the other strains. This translated to better plant shoot weights, which were similar to those of plants inoculated with WSM688 (the incumbent inoculant strain at time of testing) and 48% greater when compared to former inoculant strain CC169 (J. G. Howieson unpublished data).

The nitrogen fixation capacity (effectiveness) of *Medicago* symbioses is characterized by strong interactions between the strain of rhizobia and species of *Medicago* [13-16]. Hence, the ability to form effective symbiosis with the species recommended for inoculation is an important consideration in inoculant strain selection. WSM1115 satisfies this requirement. In greenhouse tests it formed

effective symbiosis with 16 genotypes of *Medicago* and overall produced 48% more shoot dry matter compared to plants inoculated with WSM688, the strain that it replaced (R.A. Ballard and N. Charman, unpublished data).

A limitation of strain WSM1115 is its poor persistence in moderately saline soils (e.g. where summer salinity levels exceed 10 ECe (dS/m)). Poor nodulation of regenerating pasture was first noted in 2004 during the field evaluation and domestication of the salt tolerant annual pasture legume messina (*Melilotus siculus* syn. *Melilotus messanensis*). Subsequent studies [17] confirmed that although WSM1115 was able to nodulate and form effective symbiosis with messina, it did not persist as well as other strains (e.g. SRDI554) through the summer months when salinity levels increased.

Here we present a preliminary description of the general features of *Ensifer medicae* strain WSM1115 together with its genome sequence and annotation.

Classification and features

Ensifer medicae strain WSM1115 is a motile, non-sporulating, non-encapsulated, Gram-negative rod in the order *Rhizobiales* of the class *Alphaproteobacteria*. The rod-shaped form varies in size with dimensions of approximately 0.5 µm in width and 1.0 µm in length (Figure 1A). It is fast growing, forming colonies within 3-4 days when grown on TY [18] or half strength Lupin Agar (½LA) [19] at 28°C. Colonies on ½LA are opaque, slightly domed and moderately mucoid with smooth margins (Figure 1B).

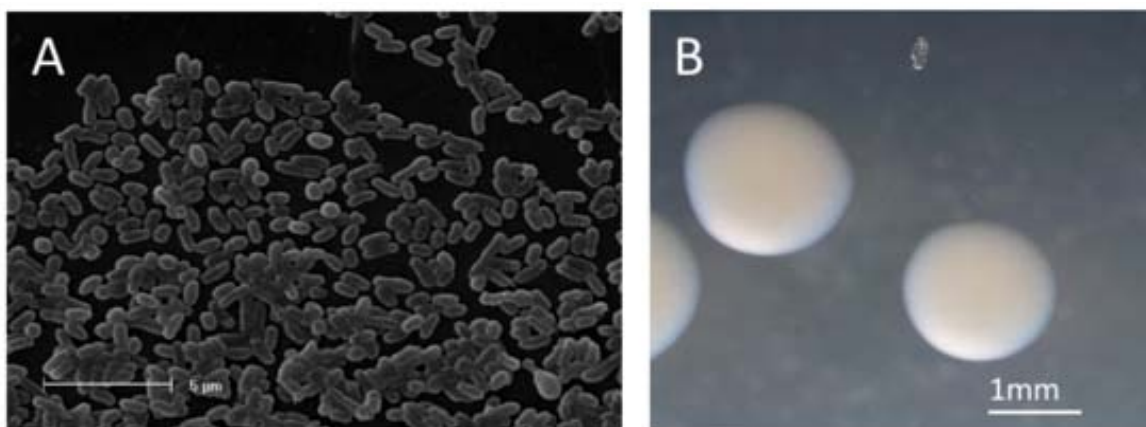


Figure 1. Images of *Ensifer medicae* strain WSM1115 using (A) scanning electron microscopy and (B) light microscopy to show the colony morphology on a solid medium.

Minimum Information about the Genome Sequence (MIGS) is provided in Table 1. Figure 2 shows the phylogenetic neighborhood of *Ensifer medicae* strain WSM1115 in a 16S rRNA gene sequence based tree. This strain has 100% sequence

identity (1,366/1,366 bp) at the 16S rRNA sequence level to the fully sequenced *Ensifer medicae* strain WSM419 [12] and 99% 16S rRNA sequence (1362/1366 bp) identity to the fully sequenced *E. meliloti* Sm1021 [36].

Table 1. Classification and general features of *Ensifer medicae* strain WSM1115 according to the MIGS recommendations [20]

MIGS ID	Property	Term	Evidence code
		Domain <i>Bacteria</i>	TAS [21]
		Phylum <i>Proteobacteria</i>	TAS [22]
		Class <i>Alphaproteobacteria</i>	TAS [23,24]
	Current classification	Order <i>Rhizobiales</i>	TAS [22,25]
		Family <i>Rhizobiaceae</i>	TAS [26,27]
		Genus <i>Ensifer</i>	TAS [28-30]
		Species <i>Ensifer medicae</i>	TAS [29]
		Strain WSM1115	
	Gram stain	Negative	IDA
	Cell shape	Rod	IDA
	Motility	Motile	IDA
	Sporulation	Non-sporulating	NAS
	Temperature range	Mesophile	NAS
	Optimum temperature	28°C	NAS
	Salinity	Non-halophile	NAS
MIGS-22	Oxygen requirement	Aerobic	IDA
	Carbon source	Varied	NAS
	Energy source	Chemoorganotroph	NAS
MIGS-6	Habitat	Soil, root nodule, on host	IDA
MIGS-15	Biotic relationship	Free living, symbiotic	IDA
MIGS-14	Pathogenicity	Non-pathogenic	IDA
	Biosafety level	1	TAS [31]
	Isolation	Root nodule	IDA
MIGS-4	Geographic location	Samothraki, Greece	IDA
MIGS-5	Time of sample collection	May, 1987	IDA
MIGS-4.1	Latitude	40.4900	IDA
MIGS-4.2	Longitude	25.6500	IDA
MIGS-4.3	Depth	<10 cm	IDA
MIGS-4.4	Altitude	325 m	IDA

Evidence codes – IDA: Inferred from Direct Assay; TAS: Traceable Author Statement (i.e., a direct report exists in the literature); NAS: Non-traceable Author Statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from the Gene Ontology project [32].

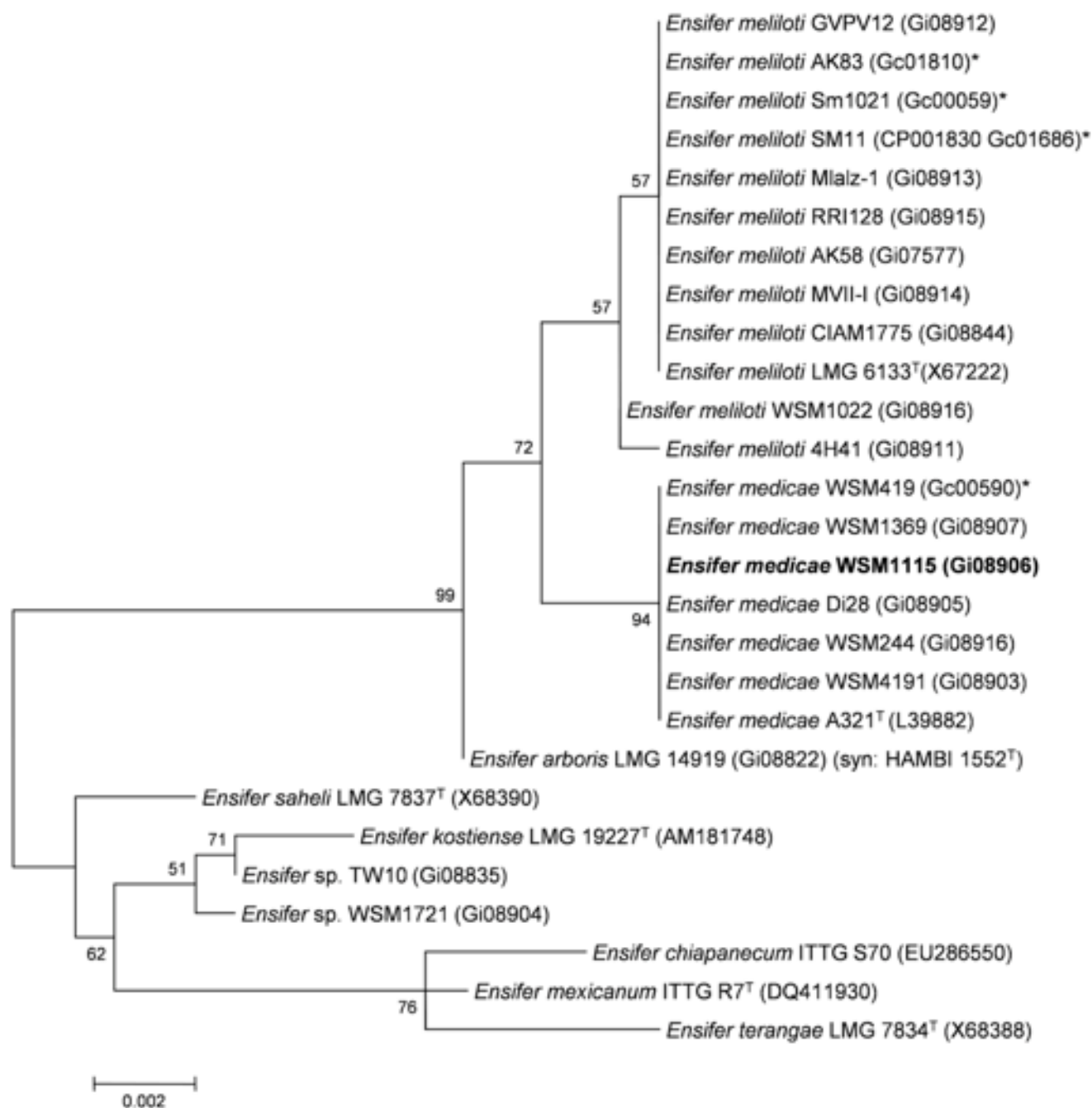


Figure 2. Phylogenetic tree showing the relationship of *Ensifer medicae* WSM1115 (shown in bold print) to other *Ensifer* spp. in the order *Rhizobiales* based on aligned sequences of the 16S rRNA gene (1,290 bp internal region). All sites were informative and there were no gap-containing sites. Phylogenetic analyses were performed using MEGA, version 5 [33]. The tree was built using the Maximum-Likelihood method with the General Time Reversible model [34]. Bootstrap analysis [35] with 500 replicates was performed to assess the support of the clusters. Type strains are indicated with a superscript T. Brackets after the strain name contain a DNA database accession number and/or a GOLD ID (beginning with the prefix G) for a sequencing project registered in GOLD [32]. Published genomes are indicated with an asterisk.

Table 2. Compatibility of *Ensifer medicae* WSM1115 with various *Medicago* and allied genera for nodulation (Nod) and N₂-fixation (Fix)

Species Name	Cultivar or line	Common Name	Growth Type	Nod	Fix	Reference
<i>M. polymorpha</i>	Santiago/Cavalier/Scimitar	Burr	Annual	+	+	IDA
<i>M. truncatula</i>	Caliph/Jester	Barrel	Annual	+	+	IDA
<i>M. murex</i>	Zodiac	Murex	Annual	+	+	IDA
<i>M. sphaerocarpus</i>	Orion	Sphere	Annual	+	+	IDA
<i>M. scutellata</i>	Sava/Silver/Essex	Snail	Annual	+	+	IDA
<i>M. rugosa</i>	Paraponto	Gama	Annual	+	+	IDA
<i>M. littoralis</i>	Herald/Harbinger	Strand	Annual	+	Poor	IDA
<i>M. orbicularis</i>	Estes	Button	Annual	+	+	[15]
<i>M. rigiduloides</i>	Accession PI 227850	Rigid	Annual	+(w)	-	[15]
<i>M. rigidula</i>	Accession PI 495552	Tifton	Annual	+(w)	-	[15]
<i>M. arabica</i>	Local ecotype	Spotted	Annual	+	+	[15]
<i>M. minima</i>	Devine	Woolly burr	Annual	+	+	[15]
<i>M. sativa</i>	SARDI Ten	Lucerne	Perennial	+	+	IDA
<i>M. lupulina</i>	'BEBLK'	Black	Perennial	+	+	[15]
<i>Melilotus siculus</i>	Accessions SA40006 & 39909	Messina	Annual	+	+	[17]
<i>Melilotus albus</i>	various accessions	Bokhara clover	Biennial	+	+	IDA

(w) indicates nodules present were white.

IDA: Inferred from Direct Assay from the Gene Ontology project [37].

Symbiotaxonomy

Ensifer medicae strain WSM1115 forms nodules (Nod+) and fixes N₂ (Fix+) with a range of annual and perennial *Medicago* species and *Melilotus* species (Table 2). Levels of N₂ fixation in combination with *Medicago littoralis* is suboptimal, that species generally forming more effective associations with strains of *Ensifer meliloti* including strain RRI128 [38]. The level of N₂ fixation with *Melilotus albus* is also noted as positive, but has been observed to vary markedly with different plant accessions.

Genome sequencing and annotation information

Genome project history

This organism was selected for sequencing on the basis of its environmental and agricultural relevance to issues in global carbon cycling, alternative energy production, and biogeochemical importance, and is part of the Community Sequencing Program at the U.S. Department of Energy, Joint Genome Institute (JGI) for projects of relevance to agency missions. The genome project is deposited in the Genomes OnLine Database [32] and a high-quality-draft genome sequence in IMG/GEBA. Sequencing, finishing and annotation were performed by the JGI. A summary of the project information is shown in Table 3.

Table 3. Genome sequencing project information for *Ensifer medicae* strain WSM1115

MIGS ID	Property	Term
MIGS-31	Finishing quality	Permanent high quality draft
MIGS-28	Libraries used	2× Illumina libraries; Std short PE & CLIP long PE
MIGS-29	Sequencing platforms	Illumina HiSeq 2000
MIGS-31.2	Sequencing coverage	530× Illumina
MIGS-30	Assemblers	with Allpaths, version 38445, Velvet 1.1.05, phrap 4.24
MIGS-32	Gene calling methods	Prodigal 1.4, GenePRIMP
	Genbank ID	AQZC01000000
	Genbank Date of Release	April 22, 2013
	GOLD ID	Gi08906
	NCBI project ID	74391
	Database: IMG-GEBA	2512875026
	Project relevance	Symbiotic N ₂ fixation, agriculture

Growth conditions and DNA isolation

Ensifer medicae strain WSM1115 was cultured to mid logarithmic phase in 60 ml of TY rich medium on a gyratory shaker at 28°C [39]. DNA was isolated from the cells using a CTAB (Cetyl trimethyl ammonium bromide) bacterial genomic DNA isolation method [40].

Genome sequencing and assembly

The genome of *Ensifer medicae* strain WSM1115 was sequenced at the Joint Genome Institute (JGI) using Illumina [41] data. An Illumina standard paired-end library with a minimum insert size of 270 bp was used to generate 23,080,558 reads totaling 3,462 Mbp and an Illumina CLIP paired-end library with an average insert size of 9,584 + 2,493 bp was used to generate 2,163,668 reads totaling 324 Mbp of Illumina data (unpublished, Feng Chen).

All general aspects of library construction and sequencing performed at the JGI can be found at the JGI user home [40]. The initial draft assembly contained 57 contigs in 11 scaffolds. The initial draft data was assembled with Allpaths, version 38445, and the consensus was computationally shredded into 10 Kbp overlapping fake reads (shreds). The Illumina draft data was also assembled with Velvet, version 1.1.05 [42], and the consensus sequences were computationally shredded into 1.5 Kbp overlapping fake reads

(shreds). The Illumina draft data was assembled again with Velvet using the shreds from the first Velvet assembly to guide the next assembly. The consensus from the second VELVET assembly was shredded into 1.5 Kbp overlapping fake reads. The fake reads from the Allpaths assembly and both Velvet assemblies and a subset of the Illumina CLIP paired-end reads were assembled using parallel phrap, version 4.24 (High Performance Software, LLC). Possible mis-assemblies were corrected with manual editing in Consed [43-45]. Gap closure was accomplished using repeat resolution software (Wei Gu, unpublished), and sequencing of bridging PCR fragments. The estimated total size of the genome is 6.9 Mbp and the final assembly is based on 3,654 Mbp of Illumina draft data, which provides an average 530× coverage of the genome.

Genome annotation

Genes were identified using Prodigal [46] as part of the Oak Ridge National Laboratory genome annotation pipeline, followed by a round of manual curation using the JGI GenePRIMP pipeline [47]. The predicted CDSs were translated and used to search the National Center for Biotechnology Information (NCBI) nonredundant database, UniProt, TIGRFam, Pfam, PRIAM, KEGG, COG, and InterPro databases. These data sources were combined to assert a product description for each

predicted protein. Non-coding genes and miscellaneous features were predicted using tRNAscan-SE [48], RNAMMer [49], Rfam [50], TMHMM [51], and SignalP [52]. Additional gene prediction analyses and functional annotation were performed within the Integrated Microbial Genomes (IMG-ER) platform [53].

Genome properties

The genome is 6,861,065 nucleotides with 61.16% GC content (Table 4) and comprised of 7 scaffolds (Figures 3a,3b,3c,3d,3e,3f and Figure 3g) From a total of 6,872 genes, 6,789 were protein encoding and 83 RNA only encoding genes. The majority of genes (76.25%) were assigned a putative function whilst the remaining genes were annotated as hypothetical. The distribution of genes into COGs functional categories is presented in Table 5.

Table 4. Genome Statistics for *Ensifer medicae* strain WSM1115

Attribute	Value	% of Total
Genome size (bp)	6,861,065	100.00
DNA coding region (bp)	5,918,651	86.26
DNA G+C content (bp)	4,196,062	61.16
Number of scaffolds	7	
Number of contigs	28	
Total gene	6,872	100.00
RNA genes	83	1.21
rRNA operons	3	0.04
Protein-coding genes	6,789	98.79
Genes with function prediction	5,240	76.25
Genes assigned to COGs	5,168	75.20
Genes assigned Pfam domains	5,424	78.93
Genes with signal peptides	571	8.31
Genes coding membrane proteins	1,483	21.58
CRISPR repeats	0	

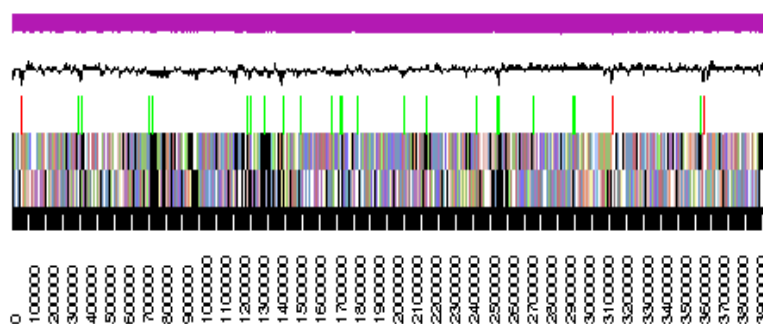


Figure 3a. Graphical maps of SinmedDRAFT_Scaffold1.2 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.

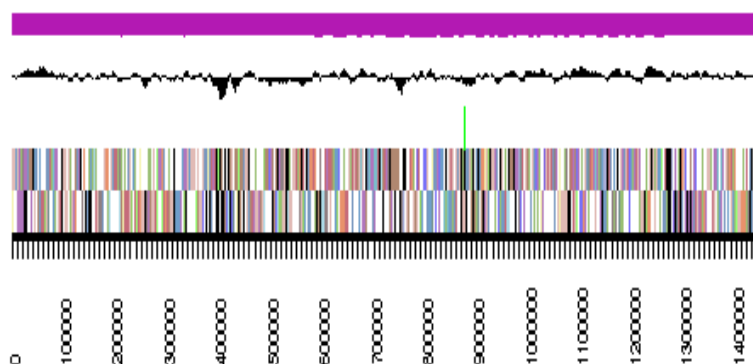


Figure 3b. Graphical maps of SinmedDRAFT_Scaffold2.1 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.

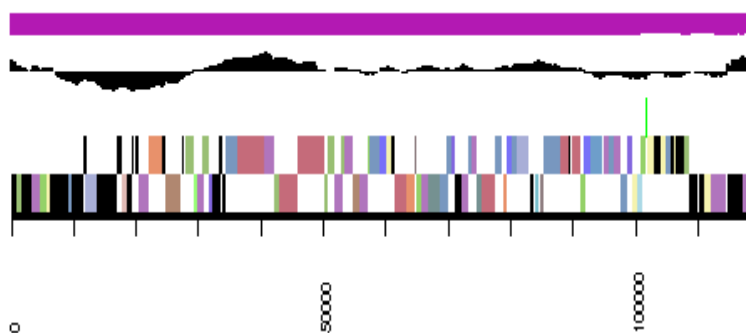


Figure 3c. Graphical maps of SinmedDRAFT_Scaffold5.3 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.

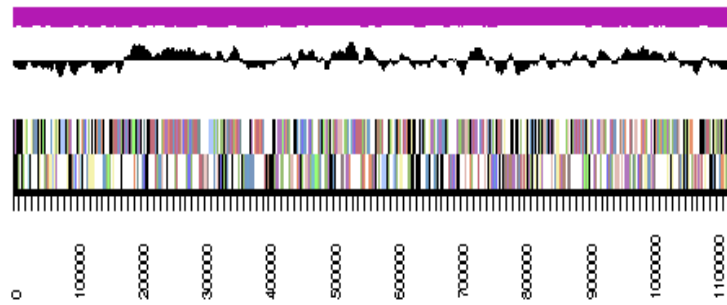


Figure 3d. Graphical maps of SinmedDRAFT_Scaffold3.7 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.



Figure 3e. Graphical maps of SinmedDRAFT_Scaffold6.5 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.

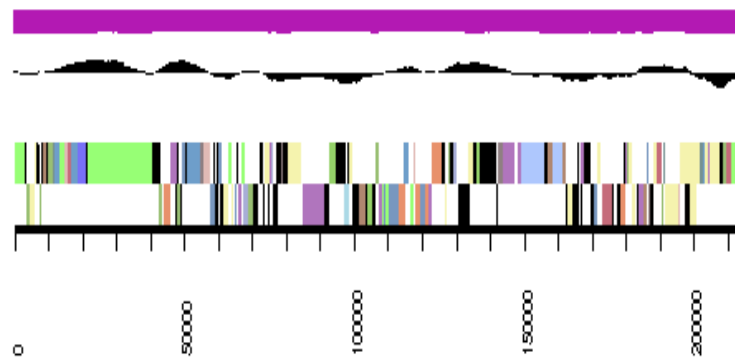


Figure 3f. Graphical maps of SinmedDRAFT_Scaffold4.6 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.



Figure 3g. Graphical maps of SinmedDRAFT_Scaffold7.4 of the *Ensifer medicae* strain WSM1115 genome sequence. From bottom to the top of each scaffold: Genes on forward strand (color by COG categories as denoted by the IMG platform), Genes on reverse strand (color by COG categories), RNA genes (tRNAs green, sRNAs red, other RNAs black), GC content, GC skew.

Table 5. Number of protein coding genes of *Ensifer medicae* strain WSM1115 associated with the general COG functional categories.

Code	Value	%age	COG Category
J	186	3.23	Translation, ribosomal structure and biogenesis
A	0	0.00	RNA processing and modification
K	527	9.16	Transcription
L	269	4.68	Replication, recombination and repair
B	3	0.05	Chromatin structure and dynamics
D	43	0.75	Cell cycle control, mitosis and meiosis
Y	0	0.00	Nuclear structure
V	55	0.96	Defense mechanisms
T	244	4.24	Signal transduction mechanisms
M	272	4.73	Cell wall/membrane biogenesis
N	68	1.18	Cell motility
Z	0	0.00	Cytoskeleton
W	1	0.02	Extracellular structures
U	112	1.95	Intracellular trafficking and secretion
O	195	3.39	Posttranslational modification, protein turnover, chaperones
C	335	5.82	Energy production conversion
G	575	10.00	Carbohydrate transport and metabolism
E	609	10.59	Amino acid transport metabolism
F	106	1.84	Nucleotide transport and metabolism
H	194	3.37	Coenzyme transport and metabolism
I	205	3.56	Lipid transport and metabolism
P	286	4.97	Inorganic ion transport and metabolism
Q	164	2.85	Secondary metabolite biosynthesis, transport and catabolism
R	726	12.62	General function prediction only
S	577	10.03	Function unknown
-	1,704	24.80	Not in COGS
-	5,752		Total

Acknowledgements

This work was performed under the auspices of the US Department of Energy's Office of Science, Biological and Environmental Research Program, and by the University of California, Lawrence Berkeley National Laboratory under contract No. DE-AC02-05CH11231, Lawrence Livermore National Laboratory under Contract No. DE-AC52-07NA27344, and Los Alamos National Laboratory under contract No. DE-AC02-06NA25396. We grate-

fully acknowledge the funding received from the Murdoch University Strategic Research Fund through the Crop and Plant Research Institute (CaPRI) and the Centre for Rhizobium Studies (CRS) at Murdoch University and the GRDC National Rhizobium Program (Project UMU63).

References

1. Small E. Alfalfa and Relatives: Evolution and Classification of Medicago. Ottawa, Canada: NRC Reserach Press; 2011.
2. Hill MJ, Donald GE. Australian Temperate Pastures Database National Pastures Improvement Coordinating Committee; 1998.
3. Peoples MB, Baldock JA. Nitrogen dynamics of pastures: nitrogen fixation inputs, the impact of legumes on soil nitrogen fertility, and the contributions of fixed nitrogen to Australian farming systems. *Aust J Exp Agric* 2001; **41**:327-346. <http://dx.doi.org/10.1071/EA99139>
4. Rome S, Brunel B, Normand P, Fernandez M, Cleyet-Marel JC. Evidence that two genomic species of *Rhizobium* are associated with *Medicago truncatula*. *Arch Microbiol* 1996; **165**:285-288. [PubMed](http://pubmed.ncbi.nlm.nih.gov/1007/s002030050328) <http://dx.doi.org/10.1007/s002030050328>
5. Rome S, Fernandez MP, Brunel B, Normand P, Cleyet-Marel JC. *Sinorhizobium medicae* sp. nov., isolated from annual *Medicago* spp. *Int J Syst Bacteriol* 1996; **46**:972-980. [PubMed](http://pubmed.ncbi.nlm.nih.gov/101099/00207713-46-4-972) <http://dx.doi.org/10.1099/00207713-46-4-972>
6. Bullard GK, Roughley RJ, Pulsford DJ. The legume inoculant industry and inoculant quality control in Australia: 1953–2003. *Aust J Exp Agric* 2005; **45**:127-140. <http://dx.doi.org/10.1071/EA03159>
7. Brockwell J, Pilka A, Holliday RA. Soil pH is a major determinant of the numbers of naturally occurring *Rhizobium meliloti* in non-cultivated soils in central New South Wales. *Aust J Exp Agric* 1991; **31**:211-219. <http://dx.doi.org/10.1071/EA9910211>
8. Denton MD, Hill CR, Bellotti WD, Coventry DR. Nodulation of *Medicago truncatula* and *Medicago polymorpha* in two pastures of contrasting soil pH and rhizobial populations. *Appl Soil Ecol* 2007; **35**:441-448. <http://dx.doi.org/10.1016/j.apsoil.2006.08.001>
9. Howieson JG, Ewing MA. Acid tolerance in the *Rhizobium meliloti*-*Medicago* symbiosis. *Aust J Agric Res* 1986; **37**:55-64. <http://dx.doi.org/10.1071/AR9860055>
10. Howieson JG, Robson AD, Abbott LK. Acid-tolerant species of *Medicago* produce root exudates at low pH which induce the expression of nodulation genes in *Rhizobium meliloti*. *Aust J Plant Physiol* 1992; **19**:287-296. <http://dx.doi.org/10.1071/PP9920287>
11. Reeve WG, Brau L, Castelli J, Garau G, Sohlenkamp C, Geiger O, Dilworth MJ, Glenn AR, Howieson JG, Tiwari RP. The *Sinorhizobium medicae* WSM419 *lpiA* gene is transcriptionally activated by FsrR and required to enhance survival in lethal acid conditions. *Microbiology* 2006; **152**:3049-3059. [PubMed](http://pubmed.ncbi.nlm.nih.gov/101099/mic.0.28764-0) <http://dx.doi.org/10.1099/mic.0.28764-0>
12. Reeve W, Chain P, O'Hara G, Ardley J, Nandesena K, Brau L, Tiwari R, Malfatti S, Kiss H, Lapidus A, et al. Complete genome sequence of the *Medicago* microsymbiont *Ensifer (Sinorhizobium) medicae* strain WSM419. *Stand Genomic Sci* 2010; **2**:77-86. [PubMed](http://pubmed.ncbi.nlm.nih.gov/104056/sigs.43526) <http://dx.doi.org/10.4056/sigs.43526>
13. Brockwell J, Hely FW. Symbiotic characteristics of *Rhizobium meliloti*: an appraisal of the systematic treatment of nodulation and nitrogen fixation interactions between hosts and rhizobia of diverse origins. *Australian Journal of Agricultural Economics* 1966; **17**:885-889.
14. Howieson JG, Nutt B, Evans P. Estimation of host-strain compatibility for symbiotic N-fixation between *Rhizobium meliloti*, several annual species of *Medicago* and *Medicago sativa*. *Plant Soil* 2000; **219**:49-55. <http://dx.doi.org/10.1023/A:1004795617375>
15. Interrante SM, Singh R, Islam MA, Stein JD, Young CA, Butler TJ. Effectiveness of *Sinorhizobium* inoculants on annual medics. *Crop Sci* 2011; **51**:2249-2255. <http://dx.doi.org/10.2135/cropsci2011.02.0076>

16. Terpolilli JJ, O'Hara GW, Tiwari RP, Dilworth MJ, Howieson JG. The model legume *Medicago truncatula* A17 is poorly matched for N₂ fixation with the sequenced microsymbiont *Sinorhizobium meliloti* 1021. *New Phytol* 2008; **179**:62-66. [PubMed](#)
<http://dx.doi.org/10.1111/j.1469-8137.2008.02464.x>
17. Bonython AL, Ballard RA, Charman N, Nichols PGH, Craig AD. New strains of rhizobia that nodulate regenerating messina (*Melilotus siculus*) plants in saline soils. *Crop Pasture Sci* 2011; **62**:427-436. <http://dx.doi.org/10.1071/CP10402>
18. Beringer JE. R factor transfer in *Rhizobium leguminosarum*. *J Gen Microbiol* 1974; **84**:188-198. [PubMed](#)
<http://dx.doi.org/10.1099/00221287-84-1-188>
19. Howieson JG, Ewing MA, D'antuono MF. Selection for acid tolerance in *Rhizobium meliloti*. *Plant Soil* 1988; **105**:179-188.
<http://dx.doi.org/10.1007/BF02376781>
20. Field D, Garrity G, Gray T, Morrison N, Selengut J, Sterk P, Tatusova T, Thomson N, Allen M, Angiuoli SV, *et al.* Towards a richer description of our complete collection of genomes and metagenomes "Minimum Information about a Genome Sequence" (MIGS) specification. *Nat Biotechnol* 2008; **26**:541-547. [PubMed](#)
<http://dx.doi.org/10.1038/nbt1360>
21. Woese CR, Kandler O, Wheelis ML. Towards a natural system of organisms: proposal for the domains *Archaea*, *Bacteria*, and *Eucarya*. *Proc Natl Acad Sci USA* 1990; **87**:4576-4579. [PubMed](#)
<http://dx.doi.org/10.1073/pnas.87.12.4576>
22. Garrity GM, Bell JA, Lilburn T. Phylum XIV. *Proteobacteria* phyl. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 2, Part B, Springer, New York, 2005, p. 1.
23. Garrity GM, Bell JA, Lilburn TG. Class I. *Alphaproteobacteria* In: Garrity GM, Brenner DJ, Krieg NR, Staley JT, editors. *Bergey's Manual of Systematic Bacteriology*. Second ed. Volume 2: New York: Springer - Verlag; 2005, p. 1.
24. Validation List No. 107. List of new names and new combinations previously effectively, but not validly, published. *Int J Syst Evol Microbiol* 2006; **56**:1-6. [PubMed](#)
<http://dx.doi.org/10.1099/ijs.0.64188-0>
25. Kuykendall LD. Order VI. *Rhizobiales* ord. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT, editors. *Bergey's Manual of Systematic Bacteriology*. Second ed: New York: Springer - Verlag; 2005. p 324.
26. Skerman VBD, McGowan V, Sneath PHA. Approved Lists of Bacterial Names. *Int J Syst Bacteriol* 1980; **30**:225-420.
<http://dx.doi.org/10.1099/00207713-30-1-225>
27. Conn HJ. Taxonomic relationships of certain non-sporeforming rods in soil. *J Bacteriol* 1938; **36**:320-321.
28. Judicial Commission of the International Committee on Systematics of P. The genus name *Sinorhizobium* Chen *et al.* 1988 is a later synonym of *Ensifer* Casida 1982 and is not conserved over the latter genus name, and the species name '*Sinorhizobium adhaerens*' is not validly published. Opinion 84. *International Journal of Systematic and Evolutionary Microbiology* 2008;58(Pt 8):1973.
29. Young JM. The genus name *Ensifer* Casida 1982 takes priority over *Sinorhizobium* Chen *et al.* 1988, and *Sinorhizobium morelense* Wang *et al.* 2002 is a later synonym of *Ensifer adhaerens* Casida 1982. Is the combination "*Sinorhizobium adhaerens*" (Casida 1982) Willems *et al.* 2003 legitimate? Request for an Opinion. *Int J Syst Evol Microbiol* 2003; **53**:2107-2110. [PubMed](#)
<http://dx.doi.org/10.1099/ijs.0.02665-0>
30. Casida LE. *Ensifer adhaerens* gen. nov., sp. nov.: a bacterial predator of bacteria in soil. *Int J Syst Bacteriol* 1982; **32**:339-345.
<http://dx.doi.org/10.1099/00207713-32-3-339>
31. Agents B. Technical rules for biological agents. TRBA (<http://www.baua.de>):466.
32. Liolios K, Mavromatis K, Tavernarakis N, Kyrpides NC. The Genomes On Line Database (GOLD) in 2007: status of genomic and metagenomic projects and their associated metadata. *Nucleic Acids Res* 2008; **36**:D475-D479. [PubMed](#)
<http://dx.doi.org/10.1093/nar/gkm884>
33. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Mol Biol Evol* 2011; **28**:2731-2739. [PubMed](#)
<http://dx.doi.org/10.1093/molbev/msr121>
34. Nei M, Kumar S. *Molecular Evolution and Phylogenetics*. New York: Oxford University Press; 2000.

35. Felsenstein J. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 1985; **39**:783-791. <http://dx.doi.org/10.2307/2408678>
36. Galibert F, Finan TM, Long SR, Puhler A, Abola P, Ampe F, Barloy-Hubler F, Barnett MJ, Becker A, Boistard P, *et al.* The composite genome of the legume symbiont *Sinorhizobium meliloti*. *Science* 2001; **293**:668-672. [PubMed](http://dx.doi.org/10.1126/science.1060966) <http://dx.doi.org/10.1126/science.1060966>
37. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, *et al.* Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet* 2000; **25**:25-29. [PubMed](http://dx.doi.org/10.1038/75556) <http://dx.doi.org/10.1038/75556>
38. Ballard RA, Slattery JF, Charman N. Host range and saprophytic competence of *Sinorhizobium meliloti* - a comparison of strains for the inoculation of lucerne, strand and disc medic. *Aust J Exp Agric* 2005; **45**:209-216. <http://dx.doi.org/10.1071/EA03126>
39. Reeve WG, Tiwari RP, Worsley PS, Dilworth MJ, Glenn AR, Howieson JG. Constructs for insertional mutagenesis, transcriptional signal localization and gene regulation studies in root nodule and other bacteria. *Microbiology* 1999; **145**:1307-1316. [PubMed](http://dx.doi.org/10.1099/13500872-145-6-1307) <http://dx.doi.org/10.1099/13500872-145-6-1307>
40. DOE Joint Genome Institute. <http://my.jgi.doe.gov/general/index.html>
41. Bennett S. Solexa Ltd. *Pharmacogenomics* 2004; **5**:433-438. [PubMed](http://dx.doi.org/10.1517/14622416.5.4.433) <http://dx.doi.org/10.1517/14622416.5.4.433>
42. Zerbino DR. Using the Velvet de novo assembler for short-read sequencing technologies. *Current Protocols in Bioinformatics* 2010;Chapter 11:Unit 11 5.
43. Ewing B, Green P. Base-calling of automated sequencer traces using phred. II. Error probabilities. *Genome Res* 1998; **8**:175-185. [PubMed](http://dx.doi.org/10.1101/gr.8.3.175) <http://dx.doi.org/10.1101/gr.8.3.175>
44. Ewing B, Hillier L, Wendl MC, Green P. Base-calling of automated sequencer traces using phred. I. Accuracy assessment. *Genome Res* 1998; **8**:175-185. [PubMed](http://dx.doi.org/10.1101/gr.8.3.175) <http://dx.doi.org/10.1101/gr.8.3.175>
45. Gordon D, Abajian C, Green P. Consed: a graphical tool for sequence finishing. *Genome Res* 1998; **8**:195-202. [PubMed](http://dx.doi.org/10.1101/gr.8.3.195) <http://dx.doi.org/10.1101/gr.8.3.195>
46. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 2010; **11**:119. [PubMed](http://dx.doi.org/10.1186/1471-2105-11-119) <http://dx.doi.org/10.1186/1471-2105-11-119>
47. Pati A, Ivanova NN, Mikhailova N, Ovchinnikova G, Hooper SD, Lykidis A, Kypides NC. GenePRIMP: a gene prediction improvement pipeline for prokaryotic genomes. *Nat Methods* 2010; **7**:455-457. [PubMed](http://dx.doi.org/10.1038/nmeth.1457) <http://dx.doi.org/10.1038/nmeth.1457>
48. Lowe TM, Eddy SR. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 1997; **25**:955-964. [PubMed](http://dx.doi.org/10.1093/nar/25.5.955) <http://dx.doi.org/10.1093/nar/25.5.955>
49. Lagesen K, Hallin P, Rodland EA, Staerfeldt HH, Rognes T, Ussery DW. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 2007; **35**:3100-3108. [PubMed](http://dx.doi.org/10.1093/nar/gkm160) <http://dx.doi.org/10.1093/nar/gkm160>
50. Griffiths-Jones S, Bateman A, Marshall M, Khanna A, Eddy SR. Rfam: an RNA family database. *Nucleic Acids Res* 2003; **31**:439-441. [PubMed](http://dx.doi.org/10.1093/nar/gkg006) <http://dx.doi.org/10.1093/nar/gkg006>
51. Krogh A, Larsson B, von Heijne G, Sonnhammer EL. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 2001; **305**:567-580. [PubMed](http://dx.doi.org/10.1006/jmbi.2000.4315) <http://dx.doi.org/10.1006/jmbi.2000.4315>
52. Bendtsen JD, Nielsen H, von Heijne G, Brunak S. Improved prediction of signal peptides: SignalP 3.0. *J Mol Biol* 2004; **340**:783-795. [PubMed](http://dx.doi.org/10.1016/j.jmb.2004.05.028) <http://dx.doi.org/10.1016/j.jmb.2004.05.028>
53. Markowitz VM, Mavromatis K, Ivanova NN, Chen IM, Chu K, Kypides NC. IMG ER: a system for microbial genome annotation expert review and curation. *Bioinformatics* 2009; **25**:2271-2278. [PubMed](http://dx.doi.org/10.1093/bioinformatics/btp393) <http://dx.doi.org/10.1093/bioinformatics/btp393>