



SHORT GENOME REPORT

Open Access

Non-contiguous finished genome sequence and description of the gliding bacterium *Flavobacterium seoulense* sp. nov.

Su-Kyoung Shin¹, Heemoon Goo², Yong-Joon Cho³, Soonsung Kwon⁴, Dongeun Yong⁴ and Hana Yi^{1,2,5*}

Abstract

Flavobacterium seoulense strain EM1321^T is the type strain of *Flavobacterium seoulense* sp. nov., a proposed novel species within the genus *Flavobacterium*. This strain is a Gram-reaction-negative, aerobic, rod-shaped bacterium isolated from stream water in Bukhansan National Park, Seoul. This organism is motile by gliding. Here, we describe the features of *Flavobacterium seoulense* EM1321^T, together with its genome sequence and annotation. The genome comprised 3,792,640 bp, with 3,230 protein-coding genes and 52 RNA genes.

Keywords: *Flavobacterium*, Gliding motility, Aerobic, *Flavobacteriaceae*

Introduction

Flavobacterium is the type genus of the family *Flavobacteriaceae* in the phylum *Bacteroidetes*. *Flavobacterium* was proposed by Bergey *et al.* [1,2] and the description was emended by Bernardet *et al.* [3]. *Flavobacterium* species have been isolated from various environments, including seawater, freshwater, river sediments, and soil [4-8]. Members of the genus *Flavobacterium* are Gram-negative, rod-shaped, yellow-pigmented, aerobic bacteria. At the time of writing, about 118 *Flavobacterium* species with validly published names have been described [9]; however, the genomes of only 14 type strains in this genus have been sequenced.

Flavobacterium seoulense sp. nov. strain EM1321^T (= KACC 18114^T = JCM 30145^T) was isolated from stream water in Bukhansan National Park, Seoul, Korea. Here, we present a summary classification and the features of *Flavobacterium seoulense* EM1321^T as well as its genome sequence and annotation.

Classification and features

Based on its 16S rRNA gene phylogeny and phenotypic characteristics, strain EM1321^T was classified as a member of the genus *Flavobacterium* (Table 1). Preliminary sequence-based identification using the 16S rRNA gene sequences in the EzTaxon database [10] indicated that strain EM1321^T was most closely related to *F. granulii* Kw05^T (GenBank accession no. AB180738) with a sequence similarity of 96.54%. This value was lower than the 98.7% 16S rRNA gene sequence similarity as a threshold recommended by Stackebrandt and Ebers [11] to delineate a new species without carrying out DNA-DNA hybridization. Subsequent phylogenetic analysis was performed using the 16S rRNA gene sequences of strain EM1321^T and related species. Sequences were aligned according to the bacterial rRNA secondary structure model using the jPHYDIT [12]. Phylogenetic trees were constructed using neighbor-joining (NJ) and maximum-likelihood (ML) methods implemented in MEGA version 5 [13]. The resultant tree topologies were evaluated by bootstrap analyses with 1,000 random samplings. Strain EM1321^T formed a monophyletic clade together with *Flavobacterium soli* [5] in both the NJ and ML trees; however, the clustering was not supported by the bootstrap analysis (Figure 1). *Flavobacterium nitratireducens* [8] was further recovered as a sister group of the monophyletic clade in the ML tree only. Based on these phylogenetic trees, *F. soli* KACC 17417^T and *F. nitratiredu-*

* Correspondence: hanayi@korea.ac.kr

¹Department of Public Health Sciences, BK21PLUS Program in Embodiment: Health-Society Interaction, Graduate School, Korea University, Seoul, Republic of Korea

²School of Biosystem and Biomedical Science, Korea University, Seoul, Republic of Korea

Full list of author information is available at the end of the article

Table 1 Classification and general features of *Flavobacterium seoulense* EM1321^T according to the MIGC recommendations [14]

MIGS ID	Property	Term	Evidence code
	Current classification	Domain <i>Bacteria</i>	TAS [15]
		Phylum <i>Bacteroidetes</i>	TAS [16,17]
		Order <i>Flavobacteriales</i>	TAS [17,18]
		Family <i>Flavobacteriaceae</i>	TAS [3,19-21]
		Genus <i>Flavobacterium</i>	TAS [1-3,22]
		Species <i>F. seoulense</i>	IDA
		Strain EM1321 ^T	IDA
	Gram stain	Negative	IDA
	Cell shape	Rod-shaped	IDA
	Motility	Gliding	IDA
	Sporulation	Non-sporulating	IDA
	Temperature range	4–35°C	IDA
	Optimum temperature	30°C	IDA
MIGS-6	Habitat	Freshwater	IDA
MIGS-6.3	Salinity	0–4%	IDA
MIGS-22	Oxygen requirement	Aerobic	IDA
	Carbon source	d-glucose, l-arabinose	IDA
MIGS-15	Biotic relationship	Free-living	IDA
MIGS-14	Pathogenicity	Non-pathogenic	NAS
MIGS-4	Geographic location	Seoul, South Korea	IDA
MIGS-5	Sample collection time	September 2013	IDA
MIGS-4.1	Latitude	37°36'52"N	IDA
MIGS-4.2	Longitude	126°59'19"E	IDA
	Isolation	Stream water	IDA

Evidence codes - IDA: Inferred from Direct Assay; TAS: Traceable Author Statement (i.e., a direct report exists in the literature); NAS: Non-traceable Author Statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from the Gene Ontology project [23]. If the evidence is IDA, the property was directly observed by one of the authors.

cens JCM 17678^T were selected as reference strains and were obtained from the corresponding culture collections for comparative study.

Strain EM1321^T was Gram-reaction negative. Cells of strain EM1321^T were rod shaped with rounded ends and motile by gliding. The cells were 1.0–1.5 µm × 0.3–0.5 µm in size (Figure 2). No flagellum was observed. The colonies were yellow in color and translucent on R2A agar medium. Growth occurred aerobically at 4–35°C, and optimal growth was observed at 30°C. The cells grew in 0–4% (w/v) NaCl. Strain EM1321^T exhibited catalase and oxidase activities. Physiological and biochemical properties were tested by using the API 20NE, API 50CH, and API ZYM systems (BioMérieux). In the API ZYM system, enzyme activity was detected for alkaline phosphatase, esterase (C4), esterase lipase (C8), leucine arylamidase, acid phosphatase, naphthol-AS-BI-phosphohydrolase, β-galactosidase, and valine arylamidase (Table 2). No activity was detected for lipase,

trypsin, α-chymotrypsin, α-galactosidase, β-glucuronidase, α-glucosidase, N-acetyl-β-glucosaminidase, cystine arylamidase, α-mannosidase, and α-fucosidase. In the API 20NE system, positive reactions were observed for nitrate reduction and negative reactions were observed for indole production, glucose fermentation, arginine dihydrolase, urease activity, and aesculin and gelatin hydrolysis. The strain assimilated d-glucose and l-arabinose, but not d-mannitol, d-mannose, d-maltose, potassium gluconate, N-acetylglucosamine, capric acid, adipic acid, malic acid, trisodium citrate, or phenylacetic acid. Acid was produced from l-arabinose, d-xylose, d-galactose, d-glucose, d-fructose, d-mannose, and d-lactose (API 50CH).

Matrix-assisted laser-desorption/ionization time-of-flight (MALDI-TOF) MS protein analysis was carried out as previously described [24]. Deposits were done from 12 isolated colonies for each strain (strain EM1321^T and reference strains). Measurements were

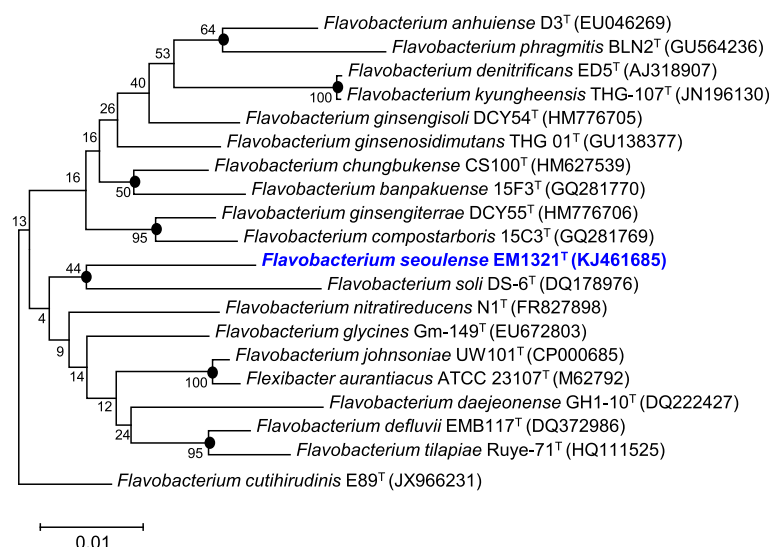


Figure 1 Phylogenetic tree highlighting the position of *Flavobacterium seoulense* EM1321^T relative to the type strains of other species within the genus *Flavobacterium*. The strains and their corresponding GenBank accession numbers of 16S rRNA genes are indicated in parentheses. The sequences were aligned using jPHYDIT and the phylogenetic inferences were obtained using neighbour-joining method with MEGA version 5 [13]. The numbers at nodes are the percentage of bootstrap values obtained by 1,000 replicates. Solid circles indicate that the corresponding nodes were also recovered in maximum-likelihood tree. Bar, 0.01 substitutions per nucleotide position.

made with a Microflex spectrometer (Bruker Daltonics, Leipzig, Germany). Spectra were recorded in the positive linear mode for the mass range of 2,000 to 20,000 Da (parameter settings: ion source 1 (IS1), 20 kV; IS2, 18.5 kV; lens, 7 kV). The time of acquisition was between 30 seconds and 1 minute per spot. The twelve EM1321^T spectra were imported into the MALDI

BioTyper software (version 2.0; Bruker) and analyzed by standard pattern matching (with default parameter settings) against 4,613 bacterial spectra including eight *Flavobacterium* species, used as reference data, in the BioTyper database. For strain EM1321^T spectrum (Figure 3), no significant score was obtained, suggesting that our isolate was not a member of the eight known species in the database. Spectrum differences with the two closely related *Flavobacterium* species are shown in Figure 4.

Genome sequencing information

Genome project history

Flavobacterium seoulense EM1321^T was selected for genome sequencing based on its phylogenetic position and its 16S rRNA similarity to other members of the genus *Flavobacterium*. The genome sequence was deposited in GenBank under accession number JNCA000000000.1. A summary of the project and the Minimum Information about a Genome Sequence (MIGS) [14] are shown in Table 3.

Growth conditions and DNA isolation

Flavobacterium seoulense EM1321^T was cultured aerobically on R2A agar medium at 30°C. Genomic DNA was extracted using the QIAamp DNA mini kit (Qiagen).

Genome sequencing and assembly

The genome of strain EM1321^T was sequenced at Chun-Lab, Inc. by using an Illumina Miseq_PE_300 system

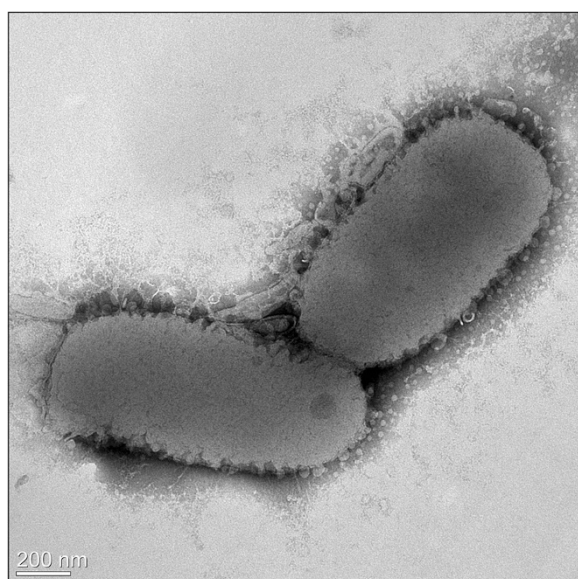


Figure 2 Transmission electron micrograph of *Flavobacterium seoulense* EM1321^T. Scale bar, 200 nm.

Table 2 Phenotypic characteristics of *Flavobacterium seoulense* EM1321^T and phylogenetically related *Flavobacterium* species

Characteristic	<i>F. seoulense</i> EM1321 ^T	<i>F. soli</i> KACC 17417 ^T	<i>F. nitratireducens</i> JCM 17678 ^T
Cell length (μm)	1.0–1.5	1.0–3.0 ^a	1.0–1.5 ^b
Oxygen requirement	Aerobic	Aerobic ^a	Aerobic ^b
Gram stain	-	- ^a	- ^b
Salt requirement	0–4%	0–2% ^a	0–1% ^b
Motility	+	+ ^a	- ^b
Spore formation	-	-	-
Production of			
Alkaline phosphatase	+	+	+
Acid phosphatase	+	+	+
Catalase	+	+	+
Oxidase	+	+	+
Nitrate reductase	+	-	+
Urease	-	-	+
α-Galactosidase	-	-	+
β-Galactosidase	+	-	-
β-Glucuronidase	-	-	-
α-Glucosidase	-	-	+
β-Glucosidase	-	+*	-
N-Acetyl-β-glucosaminidase	-	-	+
Indole	-	-	-
Esterase	+	+	+
Esterase lipase	+	+	+
Naphthol-AS-BI-phosphohydrolase	+	+	+
Leucine arylamidase	+	+	+
Cystine arylamidase	-	-	+
Valine arylamidase	+	+*	+
Utilization of			
d-glucose	+	-*	+
l-arabinose	+	-*	-
d-mannose	-	-*	+
d-mannitol	-	-	-
d-maltose	-	-*	+
G + C content (mol%)	33.25	36.9 ^a	36.3 ^b
Habitat	Freshwater	Soil ^a	Seawater ^b

+: positive result, -: negative result.

^aData from Yoon *et al.* [5].

^bData from Nupur *et al.* [8].

*Data incongruent with a previous study [5].

with 2 × 300 paired-end reads. The Illumina platform provided 166× coverage (for a total of 3,792,640 sequencing reads) of the genome. CLC Genomics Workbench (ver. 6.5.1) was used for sequence assembly and quality assessment. The final draft assembly contained 56 contigs.

Genome annotation

The genes in the assembled genome were predicted with Rapid Annotation using Subsystem Technology (RAST) server databases [25] and the gene-caller GLIMMER 3.02 [26]. The predicted ORFs were annotated by searching clusters of orthologous groups (COGs) [11] using the

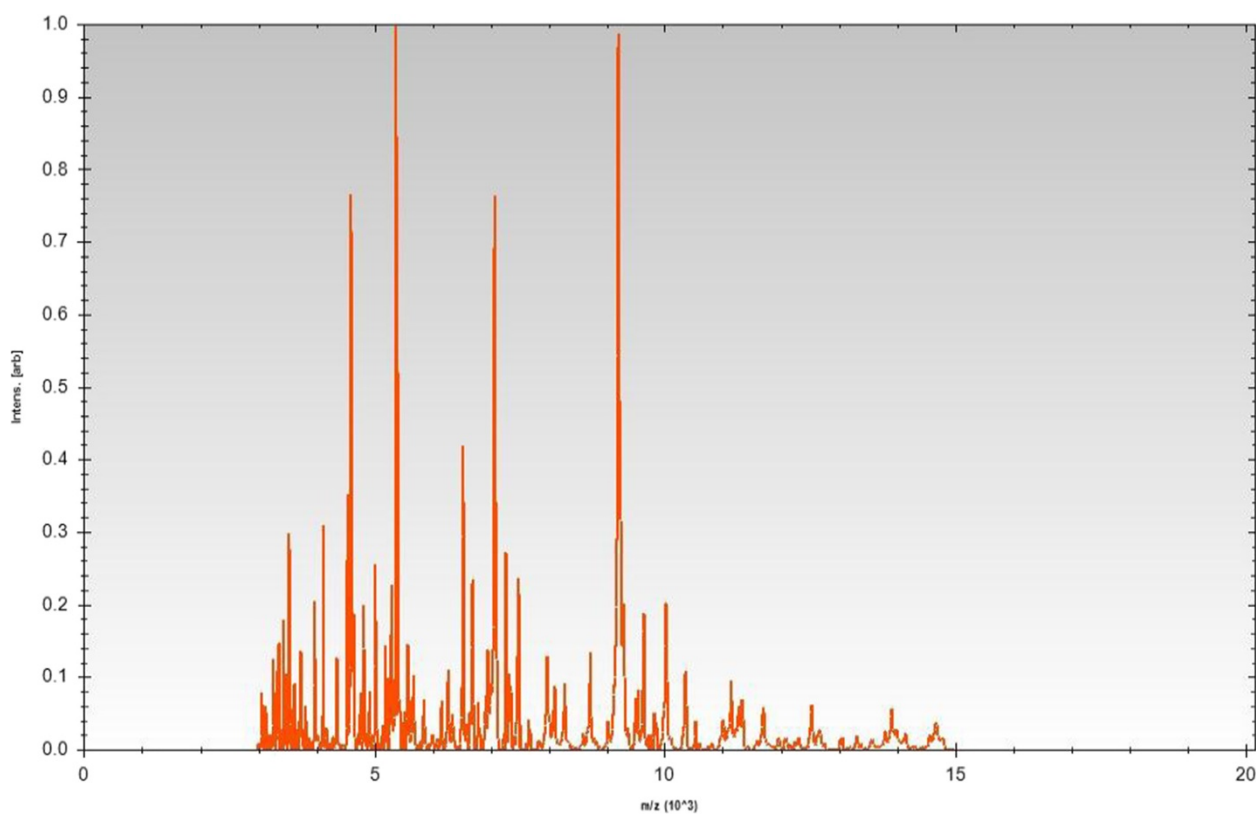


Figure 3 Reference mass spectrum from *Flavobacterium seoulense* EM1321^T. Spectra from 12 individual colonies were compared and a reference spectrum was generated.

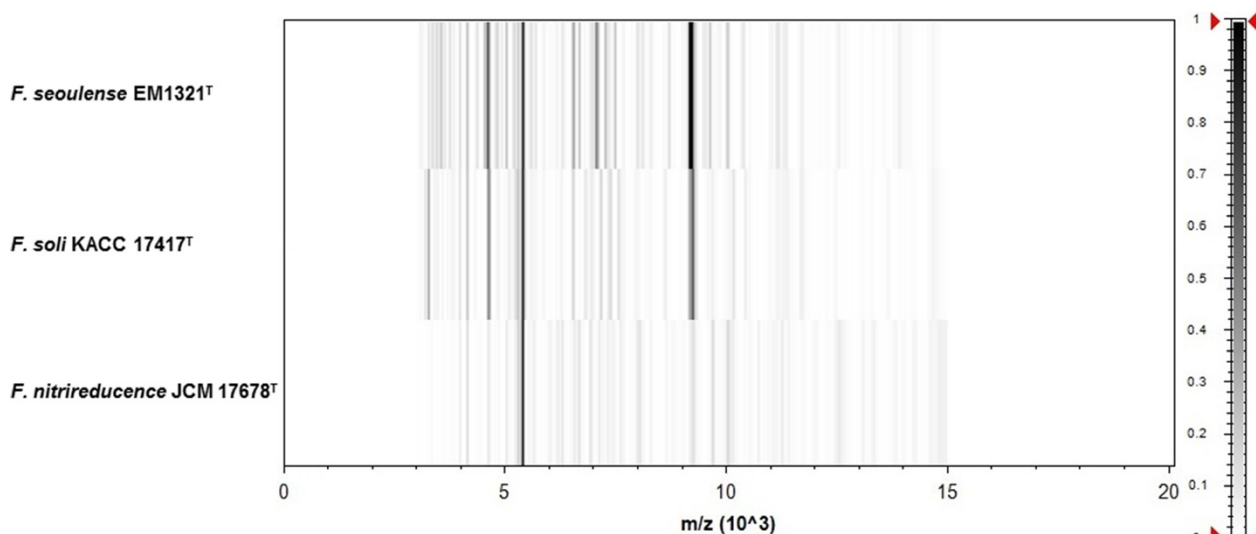


Figure 4 Gel view comparing the *Flavobacterium seoulense* EM1321^T spectrum with those of other members in the genus *Flavobacterium*. The gel view displays the raw spectra of all loaded spectrum files arranged in a pseudo-gel-like look. The x-axis records the m/z value. Peak intensity is shown as a gray-scale scheme code. The color bar and the right y-axis indicate the relation between the color of a peak and peak intensity in arbitrary units.

Table 3 Genome sequencing project information

MIGS ID	Property	Term
MIGS-31	Finishing quality	High-quality draft
MIGS-28	Libraries used	One paired-end Illumina library
MIGS-29	Sequencing platforms	Illumina MiSeq
MIGS-31.2	Fold coverage	166x
MIGS-30	Assemblers	CLCbio CLC Genomics Workbench, version 6.5.1
MIGS-32	Gene calling method	Glimmer 3.0
	Genbank ID	JNCA00000000.1
	Genbank Date of Release	2014/05/27
	BIOPROJECT	PRJNA248341
	Project relevance	Environmental, Biotechnological
MIGS-13	Source Material Identifier	KACC 18114, JCM 30145

SEED database [27]. RNAmmer 1.2 [28] and tRNAscan-SE 1.23 [29] were used to identify rRNA genes and tRNA genes, respectively. CRISPR repeats were examined using CRISPR recognition tool (CRT) [30]. CLgenomics™ 1.06 (ChunLab) was used to visualize the genomic features.

Genome properties

The genome comprised a circular chromosome with a length of 3,792,640 bp and 33.25% G + C content (Figure 5 and Table 4). It is composed of 56 contigs. Of the 3,282 predicted genes, 3,230 were protein-coding genes and 52

Table 4 Genome statistics

Attribute	Value	% of total ^a
Genome size (bp)	3,792,640	100
DNA coding region (bp)	3,386,688	89.30
G + C content (bp)	1,261,070	33.25
Total genes	3,282	100
RNA genes	52	1.58
rRNA operons	4	-
Protein-coding genes	3,230	98.42
Pseudo genes	45	1.37
Genes with function prediction	2,054	62.58
Genes assigned to COGs	2,281	69.50
Genes assigned Pfam domains	1,997	60.85
Genes with signal peptides	119	3.63
Genes with transmembrane helices	682	20.78
CRISPR repeats	0	-

^aThe total is based on either the size of the genome in base pairs or the total number of protein-coding genes in the annotated genome.

were RNA genes (2 rRNA genes and 50 tRNA genes). The sequencing coverage of rRNA operon (673x) indicated that 4 copies of rRNA operons are exist in this genome. The majority of the protein-coding genes (2,054 genes, 62.58%) were assigned putative functions, while the remaining genes were annotated as hypothetical proteins (1,176 genes, 35.83%). The properties of and statistics for the genome are summarized in Table 4. The distribution of genes into COG functional categories is presented in Table 5 and Figure 5.

Conclusions

Based on the results from phylogenetic and phenotypic analyses, we formally propose the creation of the new species *Flavobacterium seoulense* sp. nov. for strain EM1321^T. The non-contiguous genome sequence of the type strain was determined and described here.

Description of *Flavobacterium seoulense* sp. nov.

Flavobacterium seoulense (seo.ul.en'se. N.L. neut. adj., named after Seoul, Korea, the geographical origin of the type strain).

Aerobic, Gram-reaction negative. Cells are rod shaped and motile by gliding. Does not have a flagellum. The colonies are yellow in color and translucent on R2A agar medium. Grows at 4–35°C, with optimum growth at 30°C and in 0–4% (w/v) NaCl. Catalase- and oxidase-positive. Positive for alkaline phosphatase, esterase (C4), esterase lipase (C8), leucine arylamidase, acid phosphatase, naphthol-AS-BI-phosphohydrolase, β -galactosidase, and valine arylamidase. Positive for nitrate reduction, but negative for indole production, glucose fermentation, arginine dihydrolase, urease activity, and aesculin and gelatin

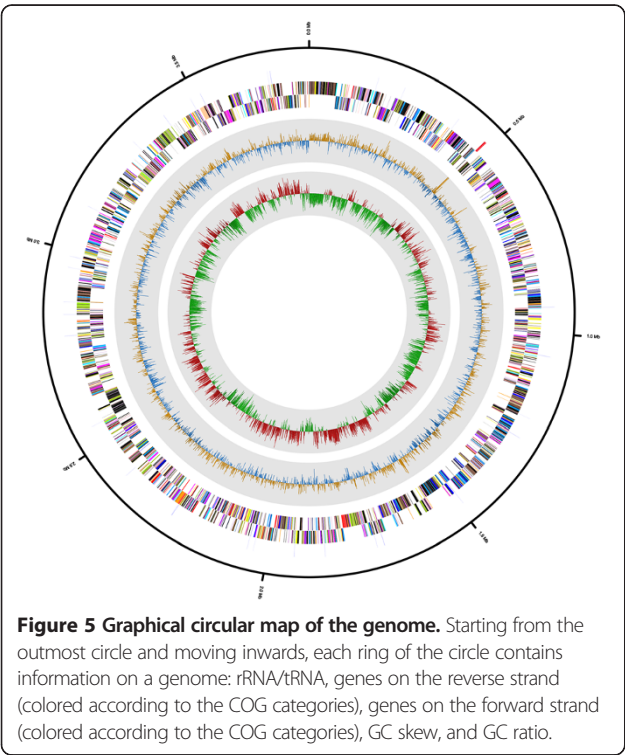


Figure 5 Graphical circular map of the genome. Starting from the outmost circle and moving inwards, each ring of the circle contains information on a genome: rRNA/tRNA, genes on the reverse strand (colored according to the COG categories), genes on the forward strand (colored according to the COG categories), GC skew, and GC ratio.

Table 5 Number of genes associated with the 25 general COG functional categories

Code	Value	% ^a	Description
J	157	4.86	Translation
A	1	0.03	RNA processing and modification
K	148	4.58	Transcription
L	123	3.81	Replication, recombination, and repair
B	0	0.00	Chromatin structure and dynamics
D	23	0.71	Cell cycle control, mitosis, and meiosis
Y	0	0.00	Nuclear structure
V	40	1.24	Defense mechanisms
T	121	3.75	Signal transduction mechanisms
M	220	6.81	Cell wall/membrane biogenesis
N	18	0.56	Cell motility
Z	0	0.00	Cytoskeleton
W	0	0.00	Extracellular structures
U	45	1.39	Intracellular trafficking and secretion
O	81	2.51	Posttranslational modification, protein turnover, and chaperones
C	122	3.78	Energy production and conversion
G	207	6.41	Carbohydrate transport and metabolism
E	170	5.26	Amino acid transport and metabolism
F	62	1.92	Nucleotide transport and metabolism
H	127	3.93	Coenzyme transport and metabolism
I	93	2.88	Lipid transport and metabolism
P	170	5.26	Inorganic ion transport and metabolism
Q	42	1.30	Secondary metabolites biosynthesis, transport, and catabolism
R	318	9.85	General function prediction only
S	196	6.07	Function unknown
-	949	29.38	Not in COGs

^aThe total is based on the total number of protein coding genes in the annotated genome.

hydrolysis. Negative for lipase, trypsin, α -chymotrypsin, α -galactosidase, β -glucuronidase, α -glucosidase, β -glucosidase, N -acetyl- β -glucosaminidase, or cystine arylamidase activity. This strain assimilated d-glucose and l-arabinose, but not d-mannitol, d-mannose, d-maltose, N -acetylglucosamine, potassium gluconate, capric acid, adipic acid, malic acid, trisodium citrate, or phenylacetic acid. Produces acid from l-arabinose, d-xylose, d-galactose, d-glucose, d-fructose, d-mannose, and d-lactose.

The G + C content of the genome is 33.25%. The 16S rRNA and genome sequences are deposited in GenBank under accession numbers KJ461685 and JNCA00000000.1, respectively. The type strain EM1321^T (= KACC 18114^T = JCM 30145^T) was isolated from stream water in Bukhansan National Park, Seoul, Korea.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SS drafted the manuscript, performed laboratory experiments, and analyzed the data. HG cultured samples and performed the electron micrograph and phylogenetic analysis. YC, SK and DY sequenced, assembled, and annotated the genome. HY organized the study and drafted the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by the Basic Science Research Programs through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT, and Future Planning (NRF-2013R1A1A3010041) and supported by a Korea University Grant.

Author details

¹Department of Public Health Sciences, BK21PLUS Program in Embodiment: Health-Society Interaction, Graduate School, Korea University, Seoul, Republic of Korea. ²School of Biosystem and Biomedical Science, Korea University, Seoul, Republic of Korea. ³Chunlab, Inc., Seoul, Republic of Korea. ⁴Department of Laboratory Medicine and Research Institute of Bacterial Resistance, Yonsei University College of Medicine, Seoul, Republic of Korea. ⁵Korea University Guro Hospital, Korea University, Seoul, Republic of Korea.

Received: 8 July 2014 Accepted: 24 November 2014

Published: 29 December 2014

References

- Bergey DH, Harrison FC, Breed RS WHB, Huntoon FM. *Bergey's Manual of Determinative Bacteriology*, Volume 4. 1st ed. Baltimore: The Williams and Wilkins Co; 1923: p. 1–442.
- Skerman VBD, McGowan V, Sneath PHA. Approved lists of bacterial names. *Int J Syst Bacteriol*. 1980; **30**:225–420.
- Bernardet JF, Segers P, Vancannet M, Berthe F, Kersters K, Vandamme P. Cutting a gordian knot: Emended classification and description of the genus *Flavobacterium*, emended description of the family *Flavobacteriaceae*, and proposal of *Flavobacterium hydatidis* nom nov (basonym, *Cytophaga aquatilis* Strohl and Tait 1978). *Int J Syst Bacteriol*. 1996; **46**:128–48.
- Kim BY, Weon HY, Cousin S, Yoo SH, Kwon SW, Go SJ, Stackebrandt E. *Flavobacterium daejeonense* sp. nov. and *Flavobacterium suncheonense* sp. nov., isolated from greenhouse soils in Korea. *Int J Syst Evol Microbiol*. 2006; **56**:1645–49.
- Yoon JH, Kang SJ, Oh TK. *Flavobacterium soli* sp. nov., isolated from soil. *Int J Syst Evol Microbiol*. 2006; **56**:997–1000.
- Tamaki H, Hanada S, Kamagata Y, Nakamura K, Nomura N, Nakano K, Matsumura M. *Flavobacterium limicola* sp. nov., a psychrophilic, organic-polymer-degrading bacterium isolated from freshwater sediments. *Int J Syst Evol Microbiol*. 2003; **53**:519–26.
- Kim JH, Kim KY, Cha CJ. *Flavobacterium chungangense* sp. nov., isolated from a freshwater lake. *Int J Syst Evol Microbiol*. 2009; **59**:1754–58.
- Nupur Bhumika V, Srinivas TN, Kumar PA. *Flavobacterium nitratreducens* sp. nov., an amylolytic bacterium of the family *Flavobacteriaceae* isolated from coastal surface seawater. *Int J Syst Evol Microbiol*. 2013; **63**:2490–96.
- NamesforLife, LLC. <http://doi.namesforlife.com/10.1601/tx.8071>.
- Kim OS, Cho YJ, Lee K, Yoon SH, Kim M, Na H, Park SC, Jeon YS, Lee JH, Yi H, Won S, Chun J. Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylogenies that represent uncultured species. *Int J Syst Evol Microbiol*. 2012; **62**:716–21.
- Stackebrandt E, Ebers J. Taxonomic parameters revisited: tarnished gold standards. *Microbiol Today*. 2006; **33**:152–55.
- Jeon YS, Chung H, Park S, Hur I, Lee JH, Chun J. JPHYDIT: a JAVA-based integrated environment for molecular phylogeny of ribosomal RNA sequences. *Bioinformatics*. 2005; **21**:3171–73.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol*. 2011; **28**:2731–39.
- Field D, Garrity G, Gray T, Morrison N, Selengut J, Sterk P, Tatusova T, Thomson N, Allen MJ, Angiuoli SV, Ashburner M, Axelrod N, Baldauf S,

- Ballard S, Boore J, Cochrane G, Cole J, Dawyndt P, De Vos P, dePamphilis C, Edwards R, Faruque N, Feldman R, Gilbert J, Gilna P, Glockner FO, Goldstein P, Guralnick R, Haft D, Hancock D, et al. **The minimum information about a genome sequence (MIGS) specification.** *Nat Biotechnol.* 2008; **26**:541–47.
15. Woese CR, Kandler O, Wheelis ML. **Towards a natural system of organisms: proposal for the domains *Archaea*, *Bacteria*, and *Eucarya*.** *Proc Natl Acad Sci USA.* 1990; **87**:4576–79.
 16. Krieg NR, Ludwig W, Euzéby J, Whitman WB, Phylum XIV. ***Bacteroidetes* phyl. nov.** In: Krieg NR, Staley JT, Brown DR, Hedlund BP, Paster BJ, Ward NL, Ludwig W, Whitman WB, editors. *Bergey's Manual of Systematic Bacteriology*, Volume 4. 2nd ed. New York: Springer; 2011: p. 25.
 17. Euzéby JP. **Validation List No. 143. List of new names and new combinations previously effectively, but not validly, published.** *Int J Syst Evol Microbiol.* 2012; **62**:1–4.
 18. Bernardet JF, Order I. ***Flavobacteriales* ord. nov.** In: Krieg NR, Staley JT, Brown DR, Hedlund BP, Paste RBJ, Ward NL, Ludwig W, Whitman WB, editors. *Bergey's Manual of Systematic Bacteriology*, Volume 4. 2nd ed. New York: Springer; 2011: p. 105.
 19. Bernardet JF, Family I. ***Flavobacteriaceae*.** In: Krieg NR, Staley JT, Brown DR, Hedlund BP, Paste RBJ, Ward NL, Ludwig W, Whitman WB, editors. *Bergey's Manual of Systematic Bacteriology*, Volume 4. 2nd ed. New York: Springer; 2011: p. 106–11.
 20. Bernardet JF, Nakagawa Y, Holmes B. **Proposed minimal standards for describing new taxa of the family *Flavobacteriaceae* and emended description of the family.** *Int J Syst Evol Microbiol.* 2002; **52**:1049–70.
 21. List Editor. **Validation List No. 41. Validation of publication of new names and new combinations previously effectively published outside the IJSEM.** *Int J Syst Bacteriol.* 1992; **42**:327–28.
 22. Holmes B, Owen RJ. **Proposal that *Flavobacterium* breve be substituted as the type species of the genus in place of *Flavobacterium aquatile* and emended description of the genus *Flavobacterium*: status of the named species of *Flavobacterium*.** Request for an Opinion. *Int J Syst Bacteriol.* 1979; **29**:416–26.
 23. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G. **Gene ontology: tool for the unification of biology. The Gene Ontology Consortium.** *Nat Genet.* 2000; **25**:25–9.
 24. Seng P, Drancourt M, Gouriet F, La Scola B, Fournier PE, Rolain JM, Raoult D. **Ongoing revolution in bacteriology: routine identification of bacteria by matrix-assisted laser desorption ionization time-of-flight mass spectrometry.** *Clin Infect Dis.* 2009; **49**:543–51.
 25. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. **The RAST Server: rapid annotations using subsystems technology.** *BMC Genomics.* 2008; **9**:75.
 26. Delcher AL, Bratke KA, Powers EC, Salzberg SL. **Identifying bacterial genes and endosymbiont DNA with Glimmer.** *Bioinformatics.* 2007; **23**:673–79.
 27. Overbeek R, Begley T, Butler RM, Choudhuri JV, Chuang HY, Cohoon M, de Crecy-Lagard V, Diaz N, Disz T, Edwards R, Fonstein M, Frank ED, Gerdes S, Glass EM, Goesmann A, Hanson A, Iwata-Reuyl D, Jensen R, Jamshidi N, Krause L, Kubal M, Larsen N, Linke B, McHardy AC, Meyer F, Neuweger H, Olsen G, Olson R, Osterman A, Portnoy V, et al. **The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes.** *Nucleic Acids Res.* 2005; **33**:5691–702.
 28. Lagesen K, Hallin P, Rodland EA, Staerfeldt HH, Rognes T, Ussery DW. **RNAmmr: consistent and rapid annotation of ribosomal RNA genes.** *Nucleic Acids Res.* 2007; **35**:3100–08.
 29. Lowe TM, Eddy SR. **tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence.** *Nucleic Acids Res.* 1997; **25**:955–64.
 30. Bland C, Ramsey TL, Sabree F, Lowe M, Brown K, Kyrpides NC, Hugenholtz P. **CRISPR recognition tool (CRT): a tool for automatic detection of clustered regularly interspaced palindromic repeats.** *BMC Bioinformatics.* 2007; **8**:209.

doi:10.1186/1944-3277-9-34

Cite this article as: Shin et al.: Non-contiguous finished genome sequence and description of the gliding bacterium *Flavobacterium seoulense* sp. nov.. *Standards in Genomic Sciences* 2014 **9**:34.

Submit your next manuscript to BioMed Central and take full advantage of:

- **Convenient online submission**
- **Thorough peer review**
- **No space constraints or color figure charges**
- **Immediate publication on acceptance**
- **Inclusion in PubMed, CAS, Scopus and Google Scholar**
- **Research which is freely available for redistribution**

Submit your manuscript at
www.biomedcentral.com/submit

